Compatibility of Alcorfix for Giardia and Cryptosporidium Antigen Testing

Brianne Couturier¹, Jason Gowans¹, and Marc Roger Couturier² ¹ARUP Laboratories, Institute for Experimental and Clinical Pathology, Salt Lake City, UT, ²University of Utah School of Medicine, Salt Lake City, UT



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Abstract

Background: Our laboratory recently converted to a single-vial stool fixative (Alcorfix) for ova & parasite testing; eliminating formalin from our laboratory and simplifying specimen processing. A limitation with this change is requiring an additional frozen stool aliquot for parasitic antigen testing. Alcorfix compatibility with antigen detection ELISAs has not been evaluated and it is unknown whether the polyvinyl alcohol (PVA) is inhibitory. We assessed and validated Alcorfix for detection of Giardia and Cryptosporidium [Crypto] antigens and whether concentrated stool sediments were also compatible.

Methods: Unpreserved stools previously tested by antigen detection ELISA for Crypto or Giardia were collected. Aliquots of the stool were preserved in Alcorfix at a 1:3 ratio and spiked with various concentrations of Crypto oocysts (n=40) or Giardia cysts (n=40). Spiked specimens of each organism were also concentrated using a Parasep concentrator tube. The pellet and the supernatant were tested for the presence of Crypto or Giardia antigen by ELISA. The pellet was tested like a fresh stool specimen (1:4 in diluent). The supernatant was directly tested without dilution. Antigen stability for the Crypto and Giardia in Alcorfix was also assessed.

Results: The analytical sensitivity was 100% (40/40) for the detection of *Giardia* and 92.5% (37/40) for Crypto. The majority of *Giardia* antigen was reactive in the pellet rather than the supernatant. Crypto antigen was also concentrated in the pellet though there was still significant reactivity in the supernatant. The stability of the antigens in Alcorfix was limited to 7 days with Crypto (vs 14 days frozen unpreserved) and 14 days for *Giardia* (same as unpreserved).

Conclusions: Alcorfix, despite containing PVA, is compatible with *Giardia* and Crypto antigen testing by ELISA. A pellet from concentrated stool is suboptimal for detecting Crypto, as low parasite burden specimens may not be detected. Furthermore, the supernatant from the concentrated stool specimen is not acceptable for antigen testing. Using a Parasep tube with Alcorfix is a compatible combination for Giardia and Crypto antigen detection only when the entire sample is filtered, pelleted, and then resuspended before testing. The remaining suspension can then be pelleted again for further microscopic examination as needed.

Methods/Results

Stability of spiked stool specimens fixed in Alcorfix by Giardia and Cryptosporidium antigen ELISAs Table 1:

Stability of *Giardia* and Crypto antigens were determined by spiking stool and preserving in Alcorfix. Aliquots were then stored at ambient, 4°C, and -20°C and tested at 4, 7 and 14 days using the TecLab Giardia II or Cryptosporidium II antigen detection ELISAs (Blacksburg, VA) per manufacturer's instructions. The optical density (OD) and qualitative interpretation were assessed. OD values \geq = 0.150 are positive for both ELISAs.

•	S1	tab	ility	of the	e Gi	iardia an-
tige	en	is	14	days	at	ambient,
4°C	. <i>.</i>	and	-20)°С.		

• Stability of the Crypto an-
tigen is 7 days at ambient, 4°C,
and -20°C.

	Gia	rdia	Crypto		
4°C	OD	OD Result		Result	
T=0	3.167	Positive	0.369	Positive	
T= 4 days	1.199	Positive	0.354	Positive	
T=7 days	3.251	Positive	0.376	Positive	
T=14 days	3.078	Positive	0.087	Negative	
Ambient					
T=0	3.167	Positive	0.369	Positive	
T=4 days	2.929	Positive	0.332	Positive	
T=7 days	3.623	Positive	0.473	Positive	
T=14 days	3.814	Positive	0.287	Positive	
-20°C					
T=0	3.167	Positive	0.369	Positive	
T=14 days	3.096	Positive	0.116	Negative	

Methods/Results

Accuracy of spiked stool specimens fixed in Alcorfix by Giardia and Cryptosporidium antigen ELISAs

Unpreserved stool, previously run on the Crypto and Giardia antigen ELISAs were collected as spiking matrices. Aliquots of stool were fixed with Alcorfix at a ratio of 1:3 and spiked with Crypto oocysts or Giardia cysts (Waterbourne Inc, New Orleans, LA) at low, medium, and high levels (defined below).

RD47

RD49

RD53

G	Siar	dia
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Giardia					
Study ID	Spiking level	OD	Study Result		
RD-04	N/A	0.066	Negative		
RD-12	N/A	0.068	Negative		
RD-48	N/A	0.069	Negative		
RD-28	N/A	0.070	Negative		
RD-08	N/A	0.073	Negative		
RD-20	N/A	0.074	Negative		
RD-36	N/A	0.074	Negative		
RD-24	N/A	0.076	Negative		
RD-16	N/A	0.077	Negative		
RD-32	N/A	0.077	Negative		
RD-44	N/A	0.095	Negative		
RD-40	N/A	0.106	Negative		
RD-63	Low	0.210	Positive		
RD-55	Low	0.231	Positive		
RD-38	Low	0.255	Positive		
RD-61	Low	0.269	Positive		
RD-14	Low	0.275	Positive		
RD-51	Low	0.300	Positive		
RD-30	Low	0.323	Positive		
RD-53	Low	0.360	Positive		
RD-65	Low	0.368	Positive		
RD-02	Low	0.388	Positive		
RD-10	Medium	0.402	Positive		
RD-46	Medium	0.418	Positive		
RD-49	Medium	0.432	Positive		
RD-42	Medium	0.452	Positive		
RD-22	Medium	0.507	Positive		
RD-58	Medium	0.508	Positive		
RD-18	Medium	0.514	Positive		
RD-05	Medium	0.559	Positive		
RD-59	Medium	0.603	Positive		
RD-67	Medium	0.606	Positive		
RD-34	Medium	0.647	Positive		
RD-26	Medium	0.709	Positive		
RD-73	High	2.427	Positive		
RD-79	High	2.930	Positive		
RD-80	High	3.322	Positive		
RD-75	High	3.446	Positive		
RD-83	High	3.623	Positive		
RD-69	High	3.654	Positive		
RD-71	High	3.727	Positive		

Low spike $= \sim 6.25 \times 10^{5}$ cysts Medium spike $= \sim 1.25 \text{ x} 10^6 \text{ cysts}$ $= 1.87 \times 10^6 \text{ cysts}$ High spike

4.070

All samples detected as expected

0.101 RD29 Negative 0.072 RD45 N/A Negative 0.098 RD28 N/A Negative 0.098 RD23 N/A Negative 0.083 RD12 N/A Negative RD26 0.094 N/A Negative 0.075 RD25 Negative N/A 0.511 RD65 Medium Positive 0.077 Negative 0.663 RD32 Medium Positive 0.616 RD33 Medium Positive RD67 Medium 0.440 Positive 0.405 RD29-P Positive Medium 0.496 RD47-P Positive Medium 0.702 Positive RD53-P Medium 0.253 RD15 Positive Medium 0.701 RD48 Medium Positive 0.230 RD51 Medium Positive 0.544 RD50 Medium Positive 0.278 RD38 Medium Positive 0.201 RD40 Medium **Positive** 0.761 RD68 Medium Positive 0.316 RD35 Medium Positive 0.588 RD70 Positive Medium 0.650 Positive RD52 Medium 0.242 RD30 Positive Low 0.359 RD9 Positive Low 0.077 RD61 Low RD49-P 0.639 Positive Low 0.081 Negative RD11 Low 0.257 RD16 Positive Low 0.986 RD46 High Positive

0.270

0.562

0.955

0.204

0.660

Positive

Positive

Positive

Positive

Positive

Crypto

0.073

0.102

0.072

Negative

Negative

Negative

Study ID | Spiking level | OD

N/A

N/A

= \sim 5.0x10 5 oocysts Low spike Medium spike = \sim 1.0x10 6 oocysts = \sim 1.5x106 oocysts High spike

High

High

High

Three discrepant specimens

RD36

RD69

RD70

RD45-P

RD10

All detected upon re-spiking

Contact: Dr. Marc Roger Couturier Ph.D., D(ABMM) **Assistant Professor of Pathology, University of Utah Medical Director, ARUP Laboratories** marc.couturier@aruplab.com



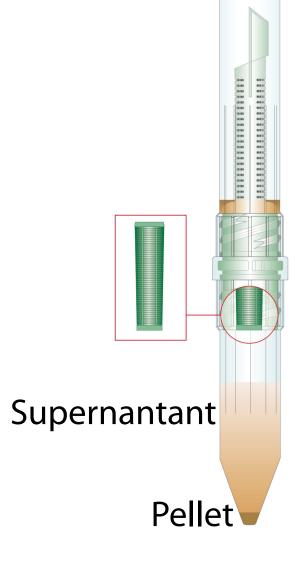
Department of Pathology

Methods/Results

TABLE 3: Pellet from concentrated stool in Alcorfix is compatible with Giardia and Cryptosporidium antigen detection

Specimens fixed in Alcorfix using Parasep Concentration Tube

- Stools were spiked with Giardia cysts or Crypto oocyts and scooped into a MIDI Parasep Concentrator tube per manufacturer's recommendations.
- Samples were concentrated per manufacture's recommendations
- A sample of the supernatant (normally discarded after concentration) and the pellet were both tested for the presence of antigen.
- The supernatant was treated as a fixed sample per ELISA protocol according to manufacturer's recommendation.
- The pellet was tested as a raw specimen and diluted 1:4 in sample diluent.



Study ID	Giardia	OD	Study Result	Study ID	Crypto	OD	Study Result
R&D1	Supernatant	0.078	Negative	R&D1	Supernatant	0.100	Negative
R&D1	Pellet	1.941	Positive	R&D1	Pellet	0.173	Positive
R&D2	Supernatant	0.729	Positive	R&D2	Supernatant	0.247	Positive
R&D2	Pellet	1.812	Positive	R&D2	Pellet	0.174	Positive
R&D3	Supernatant	0.269	Positive	R&D3	Supernatant	0.075	Negative
R&D3	Pellet	4.167	Positive	R&D3	Pellet	0.199	Positive
R&D4	Supernatant	0.206	Positive	R&D4	Supernatant	0.120	Negative
R&D4	Pellet	9.999	Positive	R&D4	Pellet	0.381	Positive
R&D5	Supernatant	0.441	Positive	R&D5	Supernatant	0.088	Negative
R&D5	Pellet	3.728	Positive	R&D5	Pellet	0.137	Negative

Giardia: Antigen was detected in both supernatant and pellet. Higher concentrations of antigen were observed in the pellet. Overall, antigen testing from the concentrated stool specimen or supernatant are compatible.

Crypto: Antigen was detected in both supernatant and pellet. R&D5 may have been spiked below the LoD. R&D2 was a watery stool and did not produce a discernible pellet which would account for the increased amount of antigen seen in the supernatant. Overall, the antigen concentrates in the pellet and it is compatible (but not optimal) for detecting Crypto antigen.

Conclusions

- The presence of PVA in Alcorfix does not significantly interfere with Giardia or Crypto antigen ELISAs from TechLab. Compatibility with other products must be investigated by individual laboratories.
- Concentrated stool specimens can be tested, but must be diluted and treated as an unpreserved specimen prior to testing.
- Frozen unpreserved stool is preferred for antigen detection, however if that is not available on submission, stool fixed in Alcorfix is also compatible*. Importantly, testing should be performed as soon as possible to ensure antigen stability, as Crypto antigen became undetectable at 7 days.

Parasep tubes kindly supplied by Apacor

*Apacor does not make claims of compatibility for Alcorfix with antigen detection assays